Programming high-order combinatorial genetics with Cas9-mediated gene drive approach for cellular engineering

Xiaoge Guo,^{1,2} (x.guo@wyss.harvard.edu), Alejandro Chavez,³ Angela Tung,¹ Christian Kaas,⁴ Aditya Kunjapur,⁵ Yi Yin,⁶ Surojit Biswas,^{1,2} Christian Kramme,^{1,2} Richie Kohman,^{1,2} Jay Shendure,⁷ James J. Collins,^{1,9,10,11,12}, **George M. Church**^{1,2,12}

¹Wyss Institute at Harvard University, Boston, MA; ²Department of Genetics, Blavatnik Institute, Harvard Medical School, Boston, MA; ³ Department of Pathology and Cell Biology, Columbia University College of Physicians and Surgeons, New York, NY; ⁴Department of Expression Technologies 2, Novo Nordisk A/S, Maaloev, Denmark; ⁵Department of Chemical and Biomolecular Engineering, University of Delaware, Newark, DE; ⁶Department of Human Genetics, University of Los Angeles, Los Angeles, CA; ⁷Department of Genome Sciences, University of Washington, Seattle, WA; ⁹Institute for Medical Engineering & Science, Massachusetts Institute of Technology, Cambridge, MA; ¹⁰Synthetic Biology Center, Massachusetts Institute of Technology, Cambridge, MA; ¹¹Department of Biological Engineering, Massachusetts Institute of Technology, Cambridge, MA; ¹²Broad Institute of MIT and Harvard, Cambridge, MA

http://arep.med.harvard.edu

Project Goals: Deciphering combinatorial genetic modifications required for host chassis optimization is key to creating robust and economical systems to realize the promise of metabolic engineering. Our goal is to develop a high-throughput microbial platform that enables rapid generation of targeted, high-order genetic alterations coupled to single-cell mapping of combinatorial genotypes to identify host genome modifications giving rise to enhanced productions of desired biomolecules.

Current apporaches to cellular engineering rely on introducing a single genetic alteration into a genome one at a time and then studying its effects of metabolic output. This "one gene at a time" approach for discovering which combination of mutations are best able to increase metabolic output is not only time-consuming and labor-intensive but also restricted in the number and order of genetic combinations that can be tested. Here, we introduce a Cas9-based gene drive framework for constructing pools of cells with combinatorial genotypes in which each cell is characterized by *N* combinations of defined genetic alteractions, along with a high-throughput method for genotype-phenotype mapping. We demonstrate the utility of our approach in rapidly swapping promoters of 19 target genes with 7 promoters representing a continuum of gene expression level and identifying all the combination of genetic manipulations that will result in high-level production of the carotenoid lycopene. Our strategy allows high-order combinatorial genetics to be explored in a high-throughput targeted manner, and greatly speed up the rate at which we are able to optimize cellular chassis to produce valuable metabolites for use in consumer, biomedical and industrial applications.

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